HIGHLIGHTS OF PRESCRIBING INFORMATION
These highlights do not include all the information needed to use HEMACORD safely and effectively. See full prescribing information for HEMACORD.

HEMACORD (hematopoietic progenitor cells, cord blood)
Injectable Suspension for Intravenous Use
Initial U.S. Approval: XXXX

WARNING: FATAL INFUSION REACTIONS, GRAFT VERSUS HOST DISEASE, ENGRAFTMENT SYNDROME, AND GRAFT FAILURE
See full prescribing information for complete boxed warning.

- Fatal infusion reactions: Monitor patients during infusion and discontinue for severe reactions. Use is contraindicated in patients with known allergy to dimethyl sulfoxide (DMSO), Dextran 40 or human serum albumin. (4, 5.1, 5.2)
- Graft-vs-host disease (GVHD): GVHD may be fatal. Administration of immunosuppressive therapy may decrease the risk of GVHD. (5.3)
- Engraftment syndrome: Engraftment syndrome may be fatal. Treat engraftment syndrome promptly with corticosteroids. (5.4)
- Graft failure: Graft failure may be fatal. Monitor patients for laboratory evidence of hematopoietic recovery. (5.5)

--------INDICATIONS AND USAGE--------
HEMACORD is an allogeneic cord blood hematopoietic progenitor cell therapy indicated for use in unrelated donor hematopoietic progenitor cell transplantation procedures in conjunction with an appropriate preparative regimen for hematopoietic and immunologic reconstitution in patients with disorders affecting the hematopoietic system that are inherited, acquired, or result from myeloablative treatment. (1)

The risk benefit assessment for an individual patient depends on the patient characteristics, including disease, stage, risk factors, and specific manifestations of the disease, on characteristics of the graft, and on other available treatments or types of hematopoietic progenitor cells. (1)

--------DOSAGE AND ADMINISTRATION--------
- Unit selection and administration of HEMACORD should be done under the direction of a physician experienced in hematopoietic progenitor cell transplantation.

- The recommended minimum dose is 2.5 x 10^7 nucleated cells/kg at cryopreservation. (2.1)
- Do not administer HEMACORD through the same tubing with other products except for normal saline. (2.3)

--------DOSAGE FORMS AND STRENGTHS--------
Each unit contains a minimum of 5 x 10^8 total nucleated cells with at least 1.25 x 10^8 viable CD34+ cells at the time of cryopreservation. The exact pre-cryopreservation nucleated cell content of each unit is provided on the container label and accompanying records. (3)

--------CONTRAINDICATIONS--------
Known sensitivity to dimethyl sulfoxide (DMSO), Dextran 40 or plasma proteins. (4)

--------WARNINGS AND PRECAUTIONS--------
- Allergic Reactions and Anaphylaxis (5.1)
- Infusion Reactions (5.2)
- Graft-versus-Host Disease (5.3)
- Engraftment Syndrome (5.4)
- Graft Failure (5.5)
- Malignancies of Donor Origin (5.6)
- Transmission of Serious Infections (5.7)
- Transmission of Rare Genetic Diseases (5.8)

--------ADVERSE REACTIONS--------
Mortality, from all causes, at 100 days post-transplant was 25%. (5, 6.1)
The most common infusion-related adverse reactions (≥5%) are hypertension, vomiting, nausea, bradycardia, and fever. (6.1)

To report SUSPECTED ADVERSE REACTIONS, contact the New York Blood Center at 1-866-767-NCBP (1-866-767-6227) and FDA at 1-800-FDA-1088 or www.fda.gov/medwatch.

--------USE IN SPECIFIC POPULATIONS--------
- Pregnancy: Based on animal data, may cause fetal harm. Use only if clearly needed. (8.1)

See 17 for PATIENT COUNSELING INFORMATION

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FULL PRESCRIBING INFORMATION: CONTENTS*

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INSTRUCTIONS FOR PREPARATION FOR INFUSION

*Sections or subsections omitted from the full prescribing information are not listed.
WARNING: FATAL INFUSION REACTIONS, GRAFT VERSUS HOST DISEASE, ENGRAFTMENT SYNDROME AND GRAFT FAILURE

Fatal infusion reactions: HEMACORD administration can result in serious, including fatal, infusion reactions. Monitor patients and discontinue HEMACORD infusion for severe reactions. Use is contraindicated in patients with known allergy to dimethyl sulfoxide (DMSO), Dextran 40 or human serum albumin. [See Contraindications (4) and Warnings and Precautions (5.1, 5.2)]

Graft-vs-host disease (GVHD): GVHD is expected after administration of HEMACORD, and may be fatal. Administration of immunosuppressive therapy may decrease the risk of GVHD. [See Warnings and Precautions (5.3)]

Engraftment syndrome: Engraftment syndrome may progress to multiorgan failure and death. Treat engraftment syndrome promptly with corticosteroids. [See Warnings and Precautions (5.4)]

Graft failure: Graft failure may be fatal. Monitor patients for laboratory evidence of hematopoietic recovery. Prior to choosing a specific unit of HEMACORD, consider testing for HLA antibodies to identify patients who are alloimmunized. [See Warnings and Precautions (5.5)]

1 INDICATIONS AND USAGE

HEMACORD is an allogeneic cord blood hematopoietic progenitor cell therapy indicated for use in unrelated donor hematopoietic progenitor stem cell transplantation procedures in conjunction with an appropriate preparative regimen for hematopoietic and immunologic reconstitution in patients with disorders affecting the hematopoietic system that are inherited, acquired, or result from myeloablative treatment.

The risk benefit assessment for an individual patient depends on the patient characteristics, including disease, stage, risk factors, and specific manifestations of the disease, on characteristics of the graft, and on other available treatments or types of hematopoietic progenitor cells.

2 DOSAGE AND ADMINISTRATION

For intravenous use only.
Do not irradiate.

2.1 Dosing

The recommended minimum dose is 2.5 x 10^7 nucleated cells/kg at cryopreservation. Multiple units may be required in order to achieve the appropriate dose.

Matching for at least 4 of 6 HLA-A antigens, HLA-B antigens, and HLA-DRB1 alleles is recommended. The HLA typing and nucleated cell content for each individual unit of HEMACORD are documented on the container label and/or in accompanying records.
2.2 Preparation for Infusion

HEMACORD should be prepared by a trained healthcare professional.

- Do not irradiate HEMACORD.
- See the appended detailed instructions for preparation of HEMACORD for infusion.
- Once prepared for infusion, HEMACORD may be stored at 4 to 25°C for up to 4 hours if DMSO is not removed, and at 4°C for up to 24 hours if DMSO is removed in a washing procedure.
- The recommended limit on DMSO administration is 1 gram per kg body weight per day. [See Warnings and Precautions (5.2)]

2.3 Administration

HEMACORD should be administered under the supervision of a qualified healthcare professional experienced in hematopoietic progenitor cell transplantation.

1. Confirm the identity of the patient for the specified unit of HEMACORD prior to administration.
2. Confirm that emergency medications are available for use in the immediate area.
3. Ensure the patient is hydrated adequately.
4. Premedicate the patient 30 to 60 minutes before the administration of HEMACORD. Premedications can include any or all of the following: antipyretic, histamine blocker, and corticosteroids.
5. Inspect the product for any abnormalities such as unusual particulates and for breaches of container integrity prior to administration. Prior to infusion, discuss all such product irregularities with the laboratory issuing the product for infusion.
6. Administer HEMACORD by intravenous infusion. Do not administer in the same tubing concurrently with products other than 0.9% Sodium Chloride, Injection (USP). HEMACORD may be filtered through a 170 to 260 micron filter designed to remove clots. Do NOT use a filter designed to remove leukocytes.
7. For adults, begin infusion of HEMACORD at 100 milliliters per hour and increase the rate as tolerated. For children, begin infusion of HEMACORD at 1 milliliter per kg per hour and increase as tolerated. The infusion rate should be reduced if the fluid load is not tolerated. The infusion should be discontinued in the event of an allergic reaction or if the patient develops a moderate to severe infusion reaction. [See Warnings and Precautions (5) and Adverse Reactions (6)]
8. Monitor the patient for adverse reactions during, and for at least six hours after, administration. Because HEMACORD contains lysed red cells that may cause renal failure, careful monitoring of urine output is also recommended.

3 DOSAGE FORMS AND STRENGTHS

Each unit of HEMACORD contains a minimum of $5.0 \times 10^8$ total nucleated cells with a minimum of $1.25 \times 10^6$ viable CD34+ cells, suspended in 10% dimethyl sulfoxide (DMSO) and 1% Dextran 40, at the time of cryopreservation.

The exact pre-cryopreservation nucleated cell content is provided on the container label and in accompanying records.
4 CONTRAINDICATIONS

HEMACORD is contraindicated in patients with known hypersensitivity to dimethyl sulfoxide (DMSO), Dextran 40 or plasma proteins. [See Description (11) and Dosage and Administration (2.2)]

5 WARNINGS AND PRECAUTIONS

5.1 Allergic Reactions and Anaphylaxis

Allergic reactions may occur with infusion of hematopoietic progenitor cells, cord blood (HPC-C), including HEMACORD. Reactions include bronchospasm, wheezing, angioedema, pruritus and hives [see Adverse Reactions (6)]. Serious hypersensitivity reactions, including anaphylaxis, also have been reported. These reactions may be due to dimethyl sulfoxide (DMSO), Dextran 40, or a plasma component of HEMACORD.

5.2 Infusion Reactions

Infusion reactions are expected to occur and include nausea, vomiting, fever, rigors or chills, flushing, dyspnea, hypoxemia, chest tightness, hypertension, tachycardia, bradycardia, dysgeusia, hematuria, and mild headache. Premedication with antipyretic, histamine antagonists, and corticosteroids may reduce the incidence and intensity of infusion reactions.

Severe reactions, including respiratory distress, severe bronchospasm, severe bradycardia with heart block or other arrhythmias, cardiac arrest, hypotension, hemolysis, elevated liver enzymes, renal compromise, encephalopathy, loss of consciousness, and seizure also may occur. Many of these reactions are related to the amount of DMSO administered. Minimizing the amount of DMSO administered may reduce the risk of such reactions, although idiosyncratic responses may occur even at DMSO doses thought to be tolerated. The actual amount of DMSO depends on the method of preparation of the product for infusion. Limiting the amount of DMSO infused to no more than 1 gm/kg/day is recommended. [See Overdosage (10)]

If infusing more than one unit of HPC-C on the same day, do not administer subsequent units until all signs and symptoms of infusion reactions from the prior unit have resolved.

Infusion reactions may begin within minutes of the start of infusion of HEMACORD, although symptoms may continue to intensify and not peak for several hours after completion of the infusion. Monitor the patient closely during this period. When a reaction occurs, discontinue the infusion and institute supportive care as needed.

5.3 Graft-versus-Host Disease (GVHD)

Acute and chronic GVHD may occur in patients who have received HEMACORD. Classic acute GVHD is manifested as fever, rash, elevated bilirubin and liver enzymes, and diarrhea. Patients transplanted with HEMACORD also should receive immunosuppressive drugs to decrease the risk of GVHD. [See Adverse Reactions (6.1)]

5.4 Engraftment Syndrome

Engraftment syndrome is manifested as unexplained fever and rash in the peri-engraftment period. Patients with engraftment syndrome also may have unexplained weight gain,
hypoxemia, and pulmonary infiltrates in the absence of fluid overload or cardiac disease. If
untreated, engraftment syndrome may progress to multiorgan failure and death. Begin treatment
with corticosteroids once engraftment syndrome is recognized in order to ameliorate the
symptoms. [See Adverse Reactions (6.1)]

5.5 Graft Failure

Primary graft failure, which may be fatal, is defined as failure to achieve an absolute neutrophil
count greater than 500/µL blood by Day 42 after transplantation. Immunologic rejection is the
primary cause of graft failure. Patients should be monitored for laboratory evidence of
hematopoietic recovery. Consider testing for HLA antibodies in order to identify patients who
are alloimmunized prior to transplantation and to assist with choosing a unit with a suitable HLA
type for the individual patient. [See Adverse Reactions (6.1)]

5.6 Malignancies of Donor Origin

Patients who have undergone HPC-C transplantation may develop post-transplant
lymphoproliferative disorder (PTLD), manifested as a lymphoma-like disease favoring non-
nodal sites. PTLD is usually fatal if not treated.

The incidence of PTLD appears to be higher in patients who have received antithymocyte
globulin. The etiology is thought to be donor lymphoid cells transformed by Epstein-Barr virus
(EBV). Serial monitoring of blood for EBV DNA may be warranted in high-risk groups.

Leukemia of donor origin also has been reported in HPC-C recipients. The natural history is
presumed to be the same as that for de novo leukemia.

5.7 Transmission of Serious Infections

Transmission of infectious disease may occur because HEMACORD is derived from human
blood. Disease may be caused by known or unknown infectious agents. Donors are screened for
increased risk of infection with human immunodeficiency virus (HIV), human T-cell
lymphotropic virus (HTLV), hepatitis B virus (HBV), hepatitis C virus (HCV), T. pallidum, T.
cruzi, West Nile Virus (WNV), transmissible spongiform encephalopathy (TSE) agents, and
vaccinia. Donors are also screened for clinical evidence of sepsis, and communicable disease
risks associated with xenotransplantation. Maternal blood samples are tested for HIV types 1
and 2, HTLV types I and II, HBV, HCV, T. pallidum, WNV, and T. cruzi. These measures do
not totally eliminate the risk of transmitting these or other transmissible infectious diseases and
disease agents. Report the occurrence of a transmitted infection to the New York Blood Center
at 1-866-767-NCBP (1-866-767-6227).

Testing is also performed for evidence of donor infection due to cytomegalovirus (CMV);
however, this is not a donor selection criterion. The result may be found on the container label
and/or in accompanying records.

5.8 Transmission of Rare Genetic Diseases

HEMACORD may transmit rare genetic diseases involving the hematopoietic system for which
donor screening and/or testing has not been performed [see Adverse Reactions (6.1)]. Cord
blood donors have been screened by family history to exclude inherited disorders of the blood
and marrow. HEMACORD has been tested to exclude donors with sickle cell anemia, and
anemias due to abnormalities in hemoglobins C, D, and E. Because of the age of the donor at the
time HPC-C collection takes place, the ability to exclude rare genetic diseases is severely
limited.

6 ADVERSE REACTIONS

Day-100 mortality from all causes was 25%.

The most common infusion-related adverse reactions (≥5%) are hypertension, vomiting, nausea,
and fever.

6.1 Clinical Trials Experience

Because clinical trials are conducted under widely varying conditions, adverse reaction rates
observed in the clinical trials of a drug cannot be directly compared to rates in the clinical trials
of another drug and may not reflect the rates observed in practice.

Infusion Reactions

The data described in Table 1 reflect exposure to 442 infusions of HPC-C manufactured by
various cord blood banks in patients treated using a total nucleated cell dose ≥2.5 × 10^7/kg on a
single-arm trial or expanded access use (The COBLT Study). The population was 60% male and
40% female of median age 5 years (range 0.05-68 years), and included patients treated for
hematologic malignancies, inherited metabolic disorders, primary immunodeficiencies, and bone
marrow failure. Preparative regimens and graft-vs-host disease prophylaxis were not
standardized. The most common infusion reactions were hypertension, vomiting, nausea and
bradycardia. Hypertension and any grades 3-4 infusion-related reactions occurred more
frequently in patients receiving HPC-C in volumes greater than 150 milliliters and in pediatric
patients. The rate of serious adverse cardiopulmonary reactions was 0.8%.

Table 1. Incidence of Infusion-Related Adverse Reactions
Occurring in ≥1% of Infusions (The COBLT Study)

<table>
<thead>
<tr>
<th>Infusion Reaction</th>
<th>Any grade</th>
<th>Grade 3-4</th>
</tr>
</thead>
<tbody>
<tr>
<td>Any reaction</td>
<td>65.4%</td>
<td>27.6%</td>
</tr>
<tr>
<td>Hypertension</td>
<td>48.0%</td>
<td>21.3%</td>
</tr>
<tr>
<td>Vomiting</td>
<td>14.5%</td>
<td>0.2%</td>
</tr>
<tr>
<td>Nausea</td>
<td>12.7%</td>
<td>5.7%</td>
</tr>
<tr>
<td>Sinus bradycardia</td>
<td>10.4%</td>
<td>0</td>
</tr>
<tr>
<td>Fever</td>
<td>5.2%</td>
<td>0.2%</td>
</tr>
<tr>
<td>Sinus tachycardia</td>
<td>4.5%</td>
<td>0.2%</td>
</tr>
<tr>
<td>Allergy</td>
<td>3.4%</td>
<td>0.2%</td>
</tr>
<tr>
<td>Hypotension</td>
<td>2.5%</td>
<td>0</td>
</tr>
<tr>
<td>Hemogloburia</td>
<td>2.1%</td>
<td>0</td>
</tr>
<tr>
<td>Hypoxia</td>
<td>2.0%</td>
<td>2.0%</td>
</tr>
</tbody>
</table>

For patients who received HEMACORD, information on infusion reactions was from voluntary
reports for 244 patients. The population included 56% males and 44% females of median age 25
years (range 0.2-73 years). Preparative regimens and graft-vs-host disease prophylaxis were not
standardized. The reactions were not graded. Eighteen per cent of patients had an infusion
reaction. The most common infusion reactions were hypertension (14%), nausea (5%), vomiting
(4%), hypoxemia (3%), dyspnea (1%), tachycardia (1%), and cough (1%). The rate of serious adverse cardiopulmonary reactions was 0.1%.

Other Adverse Reactions

For other adverse reactions, the raw clinical data from the docket was pooled for 120 adult and 1179 pediatric patients transplanted with an HPC-C total nucleated cell dose ≥2.5 x 10^7/kg. Sixty-six percent (n=862) of the 1299 patients in the docket and public data underwent transplantation as treatment for hematologic malignancy. The preparative regimens and graft-vs-host disease prophylaxis varied. The median total nucleated cell dose was 6.4 (range, 2.5-73.8) x 10^7/kg. For these patients, Day-100 mortality from all causes was 25%. Primary graft failure occurred in 16%; 42% developed grades 2-4 acute graft-vs-host disease; and 19% developed grades 3-4 acute graft-vs-host disease.

Data on other adverse reactions were available for 155 patients treated with HEMACORD at a total nucleated cell dose ≥2.5 x 10^7/kg from voluntary reports. For these patients, Day-100 mortality from all causes was 25%. Primary graft failure occurred in 15%; 43% developed grades 2-4 acute graft-vs-host disease; and 20% developed grades 3-4 acute graft-vs-host disease.

Data from published literature and from observational registries, institutional databases, and cord blood bank reviews reported to the docket for HPC-C revealed nine cases of donor cell leukemia, one case of transmission of infection, and one report of transplantation from a donor with an inheritable genetic disorder. The data are not sufficient to support reliable estimates of the incidences of these events.

In a study of 364 patients, 15% of the patients developed engraftment syndrome.

8 USE IN SPECIFIC POPULATIONS

8.1 Pregnancy

Pregnancy Category C. Animal reproduction studies have not been conducted with HEMACORD. It is also not known whether HEMACORD can cause fetal harm when administered to a pregnant woman or can affect reproduction capacity. There are no adequate and well-controlled studies in pregnant women. HEMACORD should be used during pregnancy only if the potential benefit justifies the potential risk to the fetus.

8.4 Pediatric Use

HPC-C has been used in pediatric patients with disorders affecting the hematopoietic system that are inherited, acquired, or resulted from myeloablative treatment. [See Dosage and Administration (2), Adverse Reactions (6), and Clinical Studies (14)]

8.5 Geriatric Use

Clinical studies of HPC-C did not include sufficient numbers of subjects aged 65 and over to determine whether they respond differently to HEMACORD than younger subjects. In general, administration of HEMACORD to patients over age 65 should be cautious, reflecting their greater frequency of decreased hepatic, renal, or cardiac function, and of concomitant disease or other drug therapy.
8.6 Renal Disease

HEMACORD contains Dextran 40 which is eliminated by the kidneys. The safety of HEMACORD has not been established in patients with renal insufficiency or renal failure.

10 OVERDOSE

10.1 Human Overdosage Experience

There has been no experience with overdosage of HPC-C in human clinical trials. Single doses of HEMACORD up to 5.7 x 10^7 TNC/kg have been administered. HPC-C prepared for infusion may contain dimethyl sulfoxide (DMSO). The maximum tolerated dose of DMSO has not been established, but it is customary not to exceed a DMSO dose of 1 gm/kg/day when given intravenously. Several cases of altered mental status and coma have been reported with higher doses of DMSO.

10.2 Management of Overdose

For DMSO overdosage, general supportive care is indicated. The role of other interventions to treat DMSO overdosage has not been established.

11 DESCRIPTION

HEMACORD consists of hematopoietic progenitor cells, monocytes, lymphocytes, and granulocytes from human cord blood. Blood recovered from umbilical cord and placenta is volume reduced and partially depleted of red blood cells and plasma.

The active ingredient is hematopoietic progenitor cells which express the cell surface marker CD34. The potency of cord blood is determined by measuring the numbers of total nucleated cells (TNC) and CD34+ cells, and cell viability. Each unit of HEMACORD contains a minimum of 5 x 10^8 total nucleated cells with at least 1.25 x 10^6 viable CD34+ cells at the time of cryopreservation. The cellular composition of HEMACORD depends on the composition of cells in the blood recovered from the umbilical cord and placenta of the donor. The actual nucleated cell count, the CD34+ cell count, the ABO group, and the HLA typing are listed on the container label and/or accompanying records sent with each individual unit.

HEMACORD has the following inactive ingredients: dimethyl sulfoxide (DMSO) and Dextran 40. When prepared for infusion according to instructions, the infusate contains the following inactive ingredients: Dextran 40, human serum albumin, and residual DMSO.

12 CLINICAL PHARMACOLOGY

12.1 Mechanism of Action

Hematopoietic stem/progenitor cells from HPC-C migrate to the bone marrow where they divide and mature. The mature cells are released into the bloodstream, where some circulate and others migrate to tissue sites, partially or fully restoring blood counts and function, including immune function, of blood-borne cells of marrow origin. [See Clinical Studies (14)]
In patients with enzymatic abnormalities due to certain severe types of storage disorders, mature leukocytes resulting from HPC-C transplantation may synthesize enzymes that may be able to circulate and improve cellular functions of some native tissues. However, the precise mechanism of action is unknown.

14 CLINICAL STUDIES

The effectiveness of HPC-C, as defined by hematopoietic reconstitution, was demonstrated in one single-arm prospective study, and in retrospective reviews of data from an observational database for HEMACORD and data in the dockets and public information. Sixty-six percent (n=862) of the 1299 patients in the docket and public data underwent transplantation as treatment for hematologic malignancy. Results for patients who received a total nucleated cell dose $\geq 2.5 \times 10^7$/kg are shown in Table 2. Neutrophil recovery is defined as the time from transplantation to an absolute neutrophil count more than 500 per microliter. Platelet recovery is the time to a platelet count more than 20,000 per microliter. Erythrocyte recovery is the time to a reticulocyte count greater than 30,000 per microliter. The total nucleated cell dose and degree of HLA mismatch were inversely associated with the time to neutrophil recovery in the docket data.

Table 2. Hematopoietic Recovery for Patients Transplanted with HPC-C Total Nucleated Cell (TNC) Dose $\geq 2.5 \times 10^7$/kg

<table>
<thead>
<tr>
<th>Data Source</th>
<th>The COBLT Study</th>
<th>Docket and Public Data</th>
<th>HEMACORD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Design</td>
<td>Single-arm prospective</td>
<td>Retrospective</td>
<td>Retrospective</td>
</tr>
<tr>
<td>Number of patients</td>
<td>324</td>
<td>1299</td>
<td>155</td>
</tr>
<tr>
<td>Median age (range)</td>
<td>4.6 (0.07 – 52.2) yrs</td>
<td>7.0 (&lt;1 – 65.7) yrs</td>
<td>14.5 (0.2 – 72.6) yrs</td>
</tr>
<tr>
<td>Gender</td>
<td>59% male 41% female</td>
<td>57% male 43% female</td>
<td>54% male 46% female</td>
</tr>
<tr>
<td>Median TNC Dose (range) (x $10^7$/kg)</td>
<td>6.7 (2.6 – 38.8)</td>
<td>6.4 (2.5 – 73.8)</td>
<td>4.9 (2.5 – 39.8)</td>
</tr>
<tr>
<td>Neutrophil Recovery at Day 42</td>
<td>76% (95% CI 71% – 81%)</td>
<td>77% (95% CI 75% – 79%)</td>
<td>83% (95% CI 76% – 88%)</td>
</tr>
<tr>
<td>Platelet Recovery at Day 100 (20,000/uL)</td>
<td>57% (95% CI 51% – 63%)</td>
<td>-</td>
<td>77% (95% CI 69% – 84%)</td>
</tr>
<tr>
<td>Platelet Recovery at Day 100 (50,000/uL)</td>
<td>46% (95% CI 39% – 51%)</td>
<td>45% (95% CI 42% – 48%)</td>
<td>-</td>
</tr>
<tr>
<td>Erythrocyte Recovery at Day 100</td>
<td>65% (95% CI 58% – 71%)</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Median time to Neutrophil Recovery</td>
<td>27 days</td>
<td>25 days</td>
<td>20 days</td>
</tr>
<tr>
<td>Median time to Platelet Recovery (20,000/uL)</td>
<td>90 days</td>
<td>-</td>
<td>45 days</td>
</tr>
<tr>
<td>Median time to Platelet Recovery (50,000/uL)</td>
<td>113 days</td>
<td>122 days</td>
<td>-</td>
</tr>
<tr>
<td>Median time to Erythrocyte Recovery</td>
<td>64 days</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>
16 HOW SUPPLIED/STORAGE AND HANDLING

HEMACORD is supplied as a cryopreserved cell suspension in a sealed bag containing a
minimum of $5 \times 10^8$ total nucleated cells with a minimum of $1.25 \times 10^6$ viable CD34+ cells in a
volume of 25 milliliters (NDC# 76489-001-01). The exact pre-cryopreservation nucleated cell
content is provided on the container label and accompanying records.

Store HEMACORD at or below -150°C until ready for thawing and preparation.

17 PATIENT COUNSELING INFORMATION

Discuss the following with patients receiving HEMACORD:

- Report immediately any signs and symptoms of acute infusion reactions, such as fever, chills,
fatigue, breathing problems, dizziness, nausea, vomiting, headache, or muscle aches.

- Report immediately any signs or symptoms suggestive of graft-vs-host disease, including
rash, diarrhea, or yellowing of the eyes.
1 REQUIRED EQUIPMENT, REAGENTS, AND SUPPLIES

Equipment
- Biological Safety Cabinet (BSC)
- Refrigerated blood bank centrifuge
- Plasma extractor
- Digital balance
- Tube sealer compatible with PVC plastic
- Automated cell counter
- Microscope and chamber for determining cell count and viability (optional)
- Water bath (4 liters or more)
- Canister opening tool
- Orbital Rotator

Reagents
- 5% Albumin (human), USP
- 10% Dextran 40, USP
- Bacterial culture bottles (aerobic and anaerobic)

Supplies
- Cell Wash/Infusion Bag Set (Transplant Set) (included with HEMACORD)
- Sterile Disposable Syringes: 3 mL, 30 mL and 60 mL
- Sterile tubing
- 18 gauge injection needles
- Sterile gloves
- Hemostats
- Sterile small plastic zipper-lock bags
- Alcohol prep pads
- Iodine swab sticks
- Sampling site couplers
- Tubes for cell counts, progenitor assays (optional)
- Protective cryogloves
- Transfer pack containers 300 mL
- Instructions for preparation for infusion

2 VERIFICATION OF PRODUCT IDENTITY

HEMACORD is shipped frozen in a steel canister that is contained in an insulating foam sleeve. HEMACORD must be kept at or below -150°C, either inside the container used for shipping (Dry-Shipper) or in a Liquid Nitrogen (LN2)-cooled storage device at the Transplant Center (recommended).

The bar-coded ID label of the product, affixed to the canister, is visible through the open side of the canister sleeve (Figure 1).
a. Check the HEMACORD ID label to confirm its identity with the ID of the expected product as soon as it is received.

b. Wearing protective cryogloves, transfer the HEMACORD from the Dry-Shipper to the vapor phase of a LN2 storage tank.

c. Use the canister opening tool to pry canister open at top and bottom, as shown below in Figures 2 and 3.

d. Work carefully to avoid damaging the frozen plastic product bag.

e. Check the bar-coded label on the product against your records to verify that the bar-coded and visually-readable printed number absolutely conforms to the information previously provided and the documentation included with the HEMACORD product.
f. Document this check on the “Unit Receipt Form” document received with the product.

NOTE: If there is any error or ambiguity with regard to the product ID, close the canister and keep the product at LN2 temperature. Immediately advise the staff of the New York Blood Center, Inc. (NYBC) and the transplant physician. Do not proceed until the problem is resolved. If your LN2 storage tanks have no space to store the product in its canister and insulated sleeve, add LN2 to the NYBC dry-shipper to maintain the product frozen until a completely satisfactory determination is made.

3 METHOD

3.1 Preparation of Thawing Solutions

a. Prepare the thawing solution (also called reconstitution solution) at room temperature, mixing equal volumes of 10% Dextran 40 and 5% human albumin, in a biological safety cabinet. The final concentration in the thawing solution is 5% Dextran 40 and 2.5% human albumin.
b. Attach an 18 gauge needle to a 30 cc syringe. Draw approx. 12.5 mL of 10% Dextran 40 and approx. 12.5 mL of 5% human albumin into the syringe. The contents of this syringe are to be used for diluting the cell suspension after thawing.
c. Fit 18 gauge needles to three 60 mL syringes. Draw 30 mL of 10% Dextran 40 and 30 mL of 5% human albumin into each syringe. Two of these 60 mL syringes will be used in steps “l” and “o” in section 3.4 of this procedure. The third syringe will be used in step “l” of section 3.5.
d. Alternatively, prepare the thawing solution in a 300-mL transfer bag by adding, using syringes, 150 mL 10% Dextran 40 and 150 mL 5% albumin.

3.2 Thawing HEMACORD

Wearing protective cryogloves, remove the canister with HEMACORD from the LN2 container. Keep the canister in the vapor phase, just above the surface of the LN2 for 5-10 minutes before proceeding.

Note: If two different HEMACORD products are stored in the LN2 container at the same time, open one canister at a time with the canister opening tool as described above. Carefully check the ID number on the labels attached to the canister and the product, respectively. Close the canister and leave it in the vapor phase for 5-10 min. before proceeding.

a. Open canister with the canister opening tool as described above.
b. Work carefully to avoid damaging the frozen plastic product bag. Remember that plastic at this temperature is very brittle and breaks easily.
c. Examine the bag for breaks or cracks and document this inspection on the appropriate form.
d. Remove the HEMACORD from the canister.

Caution! Do not handle the plastic bags at liquid nitrogen temperature with the tongs intended for metal canisters, as this may rip the bag. Do not allow the product or tubing to bend as it may crack.
e. Put the HEMACORD inside a zipper-locked plastic bag, let the air out and close the bag. Place the bag with the HEMACORD in a warm water bath at approximately 38°C.

f. To accelerate and homogenize thawing, carefully agitate the product bag in the water and gently knead its contents.

g. Inspect and watch for leakage. If product leaks out into the zipper-locked bag, find the site of the leak in the freezing bag and position the bag so as to prevent further escape of product. While maintaining the bag in that position, finish thawing the product. (See Section 5 for emergency product recovery in the event of a container failure.)

h. As soon as the bag’s contents become slushy, remove the bag from the water bath and place it inside a biological safety cabinet.

3.3 Connecting the Freezing Bag to the Transplant Set

The procedure to restore the osmolarity of the HPC-C cell suspension, and either remove the supernatant with DMSO or simply dilute the thawed HEMACORD, is assisted by a sterile, empty, transplant bag set designed with two spike tubes to drain both compartments of the freezing bag (see Figure 4: “Cell Wash/Infusion Bag Set”). The Cell Wash/Infusion Bag Set is included with this shipment.

*Note: The following procedure must be done in a biological safety cabinet.*

![Figure 4. Cell Wash/Infusion Bag Set](image-url)
a. Close all clamps on the Cell Wash/Infusion Bag Set.
b. Remove the HEMACORD freezing bag from the zipper-locked bag.
c. Disinfect the covers of both ports of the freezing bag with iodine.
d. Using a clean and disinfected scissors, cut off the hermetically sealed covers of the freezing bag’s spike ports (Figure 5).

e. Disinfect the cut surfaces of the spike port area of the freezing bag using iodine swab sticks (Figure 6).
f. Insert the spikes of the Cell Wash/Infusion Bag Set into the ports of the freezing bag.
g. Label the transplant bag (shown in Figure 4) with HEMACORD ID number and the name of the recipient (or label according to local standard operating procedure).

3.4 Reconstitute (dilute) the thawed HEMACORD

The amount of thawing solution used for HEMACORD is at least 5 times the volume of the frozen product including the cryoprotectant. For example, 25 mL products are diluted to 170 mL total, and thus, a volume of 145 mL of thawing solution is required to make the final volume of 170 mL in a transplant bag.
Add first a volume of thawing solution equal to the volume of thawed HEMACORD (1:1 ratio).

b. Attach the 30 cc syringe with the 25 mL thawing solution to the female luer lock of the Cell Wash/Infusion Bag Set.

c. Open PC-1, PC-2 and PC-3 (see Figure 4 above) and slowly introduce half (~12.5 mL) of the thawing solution to the 25 mL product in the freezing bag while mixing the fluids in the bag using an orbital rotator.

d. Rinse well to remove cells from the bag’s ports.

e. Close PC-3. Open PC-4 and drain the contents from the freezing bag into the transplant bag.


g. Slowly add the remaining thawing solution (~12.5 mL) to the transplant bag while mixing the fluids in the bag.

h. Close PC-3.

i. Allow approx. 5 minutes for equilibration.

j. Open PC-1 and PC-2. Pass the diluted HEMACORD back and forth between the transplant bag and the freezing bag in order to more completely wash all cells out of the freezing bag and into the transplant bag.


l. Attach a syringe with 60 mL thawing solution to the luer lock.

m. Open PC-3.

n. Transfer the 60 mL solution to the diluted HEMACORD in the transplant bag while mixing the fluids in the bag.

o. Repeat with a second 60 mL syringe. The final volume should be approx. 170 mL (50 mL diluted HEMACORD with 120 mL thawing solution).


q. Pass the reconstituted HEMACORD back and forth between the transplant bag and the freezing bag in order to wash all cells completely out of the freezing bag and into the transplant bag.

r. Close PC-4.

s. Seal the Cell Wash/Infusion Bag Set tubing between PC-4 and IP-1.

t. Cut through seal to separate the transplant bag from the freezing bag.

u. Discard the freezing bag, the luer lock, and the connecting tubing.

v. The reconstituted product can be used for infusion into a patient with or without the additional step of DMSO removal (Section 3.5 below).

w. The recommended expiration time of the reconstituted unwashed HEMACORD is four hours either at room temperature or at 4°C from the time of thaw.

x. Remove a small volume from the reconstituted product for Complete Blood Counts (CBC), CFU, CD34+ counts, viability, and sterility samples (bacterial and fungal cultures) as per transplant center procedures.

NOTE: If more than four hours elapse between thawing and infusion, an aliquot of the product should be removed and tested immediately before administration to the patient to determine the cell viability of the infused product.

y. Call the Transplant Unit to advise them that the product is ready for infusion if you do not intend to remove the cryoprotectant.

3.5 Removing the Cryoprotectant (Washing)

a. Place the transplant bag and the transfer bag in a centrifuge cup.
b. Fully support the transplant bag with inserts to prevent formation of creases during centrifugation (as shown in Figure 7 below).

c. Close SC-1 securely.
d. Centrifuge at 400 x G for 20 minutes at 10°C.
e. After centrifugation, carefully remove the bags from the centrifuge bucket without disturbing the cellular pellet in the transplant bag.
f. Place the transplant bag in the plasma extractor.
g. Using SC-1 to adjust the flow, very slowly transfer approximately 2/3 of the supernatant (Supernatant-1) to the transfer bag avoiding the passage of cells.
h. Leave approximately 1/3 of supernatant with the cells (white and sedimented red cells in the diagram above). If you detect passage of cells to the transfer bag, return the contents to the transplant bag, resuspend the cells, and repeat the centrifugation or centrifuge only the Supernatant-1 bag (as described below).
i. Empty the tubing between the bags by pushing air from the transfer bag to the transplant bag.
j. Close SC-1.
k. Seal the tubing between the bags close to the transplant bag. Cut through the seal and disconnect the transfer bag with the Supernatant-1 from the transplant bag with the cellular pellet (product).
l. Resuspend the cellular pellet by slowly adding (with a syringe) 25-50 mL of the thawing solution through the IP-1, with continuous mixing. The resuspended cells constitute the Sediment-1 (the graft).
m. The weight of the empty transplant bag is 23.6 g if cut and sealed as shown below (Figure 8). Calculate the weight of the Sediment-1 by weighing the filled transplant bag and subtracting 23.6 g.
n. Remove a small volume from the Sediment-1 for cell count, viability determination, and sterility (bacterial and fungal cultures).
o. The recommended expiration time for HEMACORD after the removal of the cryoprotectant is 24 hours from the date and time of thaw. Store the product at 4°C in a blood storage refrigerator until the product is used.
p. Inspect the supernatant for escaped cells, even if there is no appearance of escape.
q. Express 10 mL from the Supernatant-1 bag into a conical centrifuge tube (accurate volume will help the accuracy of estimations).
r. Centrifuge at 600 x G for 10 minutes.
s. Carefully aspirate 9.5 mL of supernatant without disturbing the (possible) cell pellet in the tip of the tube.
t. Resuspend the cell pellet thoroughly in the 0.5 mL of supernatant and load into a cell-counting chamber.
u. Count the nucleated cells per microliter and calculate the total number of cells in the remaining volume of Supernatant-1.
v. Determine the number of nucleated cells in Supernatant-1 per kg of patient’s weight. The transplant physician may decide whether to add these cells to Sediment-1 cells (the graft) in cases where the Sediment-1 cell dose is low or borderline.
w. If collection of escaped cells from the bag containing Supernatant-1 is desired:
   1. Centrifuge the Supernatant-1 bag at 400 X G for 20 minutes at 10°C to sediment the cells.
   2. In a laminar flow hood, connect a 300 mL transfer bag to the bag containing the centrifuged product.
   3. Position the bag in the plasma extractor and express the new supernatant (Supernatant-2) into the transfer bag, leaving the sedimanted cells (Sediment-2) in the original bag.
   4. Seal the tubing between the bags, cut through the seal, and disconnect the transfer bag with the Supernatant-2 from the original bag with the Sediment-2.
   5. Resuspend the Sediment-2 in 10-15 mL thawing solution, using a syringe and mixing gently. The transplant physician may modify the volume for injection if preferred. If volume modification is desired, resuspend the cellular pellet to the final volume by injecting with thawing solution.
   6. Weigh the Supernatant-2 bag and the Sediment-2 bag, and calculate the volumes by subtracting the weight of the empty bags similarly sealed.
   7. Remove a small volume from the Sediment-2 for cell count, viability determination, and sterility testing.
x. Bring the transplant bag (Sediment-1 bag) to the Transplant Unit, even if the second bag (Sediment-2 bag) is being prepared; the second bag can be infused separately afterwards.

4. ADMINISTRATIVE REQUIREMENTS

a. Prepare a report on the procedure. Note the condition of the HEMACORD bag, including whether and at what stage leaks or cracks were detected. Record the following:
   - HEMACORD ID number
   - Date of receipt of the HEMACORD
   - Liquid Nitrogen Storage conditions in your facility
   - Date of thawing
   - Volume of the final product
   - Total nucleated cell (TNC) count, CD34+ content
   - Viability of the cells recovered (TNC or CD34+ cells) and the method used
   - Results of bacterial and fungal cultures

b. E-mail or fax a copy of the report to the New York Blood Center, Inc.
   - Email: ncbp@nybloodcenter.org
   - Fax: (718) 707-3747

c. Keep a copy for your processing lab records.

d. Return the dry shipper to the New York Blood Center, Inc. The return address is:
   - New York Blood Center, Inc.
   - National Cord Blood Program
   - 45-01 Vernon Blvd.
   - Long Island City, NY 11101
   - Ph: (718) 706-5211
   - Fax: (718) 707-3741

5. EMERGENCY PRODUCT RECOVERY IN THE EVENT OF A CONTAINER FAILURE

a. To prevent accidental fracture, handle the HEMACORD bags with extreme caution when removing them from the protective metal cassettes, during inspection, and during the thawing process.

b. Perform the thawing process in a controlled laboratory environment that provides appropriate equipment and supplies for post-thaw sampling and/or bag rescue, as well as dedicated space and personnel for product preparation.

c. To mitigate the extreme temperature change from storage at -196°C (Liquid Nitrogen phase) to thawing at 38°C, and possible sudden vaporization of liquid nitrogen in recess of the bag or tubing, hold the HEMACORD bag in the vapor phase for a few minutes following removal from the liquid phase of nitrogen before removal for thawing.

d. To prevent an accidental drop onto the floor, handle HEMACORD bags over a flat surface, such as a table.

e. Place HEMACORD bags in individual sterile zipper-locked bags prior to thawing to facilitate salvage of the product and to reduce contamination in case of an unanticipated problem.

f. If the HEMACORD bag is obviously fractured upon removal from cold storage, or if it fractures during the thawing process, please notify the Processing Laboratory of the National Cord Blood Program at the New York Blood Center [phone number: 718-706-5211 or 1-866-767-NCBP (1-866-767-6227)] as soon as possible. Notify the transplant physician and the laboratory director immediately.
g. It is the transplant physician’s (or designee’s) responsibility to determine whether the HEMACORD product will be used or discarded and whether additional product(s) are to be requested for infusion.

h. If the transplant physician (or designee) determines that the product in a ruptured bag should be used, the HEMACORD product may be recovered as follows:

1. Place the ruptured bag into the sterile zipper-locked plastic bag to prevent further loss and/or contamination of the product during the thawing process.

2. Thaw the product according to the Section 3 above. Small leaks or tears of the ruptured bag can be blocked off with hemostat clips.

3. Withdraw the thawed product from the freezing bag and any product from the zipper-locked bag into one or more 60 mL syringe(s) with sterile tubing attached.

4. Inside a biological safety cabinet, transfer the product into a new bag using a sterile syringe. (This new bag could be either the sterile transplant bag that is provided with the HEMACORD product or a bag of a stocked salvage kit that should be readily available in the thawing laboratory for use in these situations.)

5. Save an aliquot of the product to send for gram stain and bacterial and fungal cultures.

6. Dilute (reconstitute) the thawed HEMACORD and remove the cryoprotectant according to the procedure described above or administer the diluted product to the patient as per transplant physician’s instructions.

7. It is the transplant physician’s (or designee’s) responsibility to determine whether to treat the patient with broad-spectrum antibiotic coverage and the necessity for an infectious disease consultation.

8. If possible, place the ruptured bag (with or without the product) into a biohazard bag and save for reference when notifying the National Cord Blood Program at the New York Blood Center. This staff will notify the manufacturer and provide information for returning the bag to the manufacturer for evaluation.


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